

Phytochemical Analysis of the Essential Oils From Aerial Parts of Four *Scutellaria* “Skullcap” Species Cultivated in South Alabama: *Scutellaria baicalensis* Georgi, *S. Barbata* D. Don, *S. Incana* Biehler, and *S. Lateriflora* L

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Abstract

Scutellaria (skullcap) are important medicinal plants. *Scutellaria baicalensis* and *S. barbata* have been used in Chinese traditional medicine, while *S. incana* and *S. lateriflora* were used as herbal medicines by Native Americans. In this work, the essential oils of *Scutellaria baicalensis* Georgi, *Scutellaria barbata* D. Don, *Scutellaria incana* Biehler, and *Scutellaria lateriflora* L. were obtained from plants cultivated in south Alabama and analyzed by gas chromatographic techniques, including chiral gas chromatography. The most abundant components in the *Scutellaria* essential oils were 1-octen-3-ol (31.2% in *S. incana*), linalool (6.8% in *S. incana*), thymol (7.7% in *S. barbata*), carvacrol (9.3% in *S. baicalensis*), (E)-β caryophyllene (11.6% in *S. baicalensis*), germacrene D (39.3% in *S. baicalensis*), (E)-nerolidol (10.5% in *S. incana*), palmitic acid (15.6% in *S. barbata*), phytol (19.7% in *S. incana*), and linolenic acid (8.0% in *S. barbata*). These analyses of the essential oil compositions and enantiomeric ratios of predominant aromatic molecules add to our understanding of the medicinal phytochemistry of the genus *Scutellaria*.

Keywords

essential oil, ethnobotany, volatile phytochemistry, enantiomeric distribution, skullcap, Alabama, Lamiaceae, gas chromatography

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Scutellaria baicalensis Georgi, *Scutellaria barbata* D. Don, *Scutellaria incana* Biehler, and *Scutellaria lateriflora* L. were cultivated in southern Alabama where two of the species, *S. incana* and *S. lateriflora*, are natives. These herbs, commonly and collectively referred to as ‘skullcap’, belong to the mint family (Lamiaceae) and are found across the globe in temperate regions on almost every continent.

This is part of a larger ongoing study of essential oils from the medicinal plants traditionally used by Native American tribes of the United States, in particular the Cherokee of the southeastern United States.^{1–14} The Cherokee have a rich tradition of using plants as medicines including many of the aromatic species native to the southeastern U.S. that are rich in essential oils and aromatic compounds. Many ethnobotanical accounts have been given in the anthropologies of the Cherokee, and these works were studied to determine which plants to examine in further detail.^{15–22} Two of the species of *Scutellaria* studied here were used by the Cherokee (*S. incana* and *S. lateriflora*) for at least hundreds of years, while the other 2

have been used in traditional Asian phytotherapies for several millennia (*S. baicalensis* and *S. barbata*).²³ All four were easily cultivated in the climate of south Alabama, United States, and are growing as perennials as they are being observed sprouting new growth during the spring of their second year.

The genus *Scutellaria* encompasses over 470 species with more than 70 of them being used traditionally as plant medicines.^{24,25} *Scutellaria incana*, or ‘downy skullcap,’ is a perennial

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herb preferring medium-wet to dry conditions and full sun to partial shade. It is native to the central and eastern United States, found from Michigan and New York in the north, to the southern tip of Florida in the south, and can be found in bloom between July and September.^{26,27} *Scutellaria lateriflora*, 'blue skullcap, mad-dog skullcap, or American skullcap', is also native to the United States and is the most common species of *Scutellaria* found in North America. It can be found in zones 2-7 blooming in July through September across most of the North American continent, from Florida to Canada and California to New York, preferring wetlands and full sun.^{28,29} The other 2 species, *S. barbata* and *S. baicalensis*, are also perennial herbs, both native to China, with *S. barbata*, 'barbed skullcap,' being prominent throughout southeast Asia as well as north into the mountains of Nepal. Wetlands and the edges of streams and rice fields are common places to find *S. barbata* and it usually flowers from April to June.^{30,31} Commonly called 'baikal' and found across eastern Europe and Russia and across Mongolia and China and in Korea and Japan, *S. baicalensis* is the fourth herb studied here. It can be found blooming usually between July and August along roadsides and in disturbed and cultivated areas as a weed, as well as covering grassy hillsides in the wild.³²

Many tribes of Native Americans were known to use *Scutellaria* as a medicinal plant. The Cherokee used *Scutellaria incana* roots as an abortifacient and for diarrhea treatment. They also used a decoction of the plant to soothe the nerves. They made a compound from the plant which was used topically to aid in the expelling of afterbirth as well as a kidney medicine and for breast pains. This could refer either to mastitis or breast tumors as these *Scutellaria* species have been studied for their ability to act as anti-inflammatory agents and have potent cytotoxic anti-tumor effects.³³ The roots of *S. lateriflora* were used in an infusion to calm diarrhea and to encourage menstruation. A decoction of the roots was taken as an emetic and to expel afterbirth. It was also made into a compound to be applied to help expel afterbirth. According to Mooney, in his 1885 research, the *Scutellaria* species were used to bring about menstruation and to counteract the perceived negative effects of eating food cooked by a woman who was menstruating.¹⁷ The Iroquois Native Americans of New York and Ontario used an infusion of dried, ground roots of *S. lateriflora* to prevent smallpox and for throat problems.²² The Miwok of modern-day central California used multiple species of *Scutellaria* in a decoction for cold and cough remedy and as an eye wash..³⁴

In traditional Chinese medicine (TCM) *S. baicalensis* is known as 'huang qin' and is used to treat inflammations and fevers.³² Russian research into phytomedicinals has shown that *S. baicalensis* has antibacterial, anthelmintic, anti-schlerotic, astringent, digestive stimulant, hemostatic, and anti-allergen properties.³⁵ The common name in TCM for *S. barbata* is 'ban zhi lian' and it is traditionally used as both an internal and external anti-inflammatory. Both *S. barbata* and *S. baicalensis* have also been used as anxiolytics and sedatives.^{30-32,36}

Table 1. Essential Oil Yields After Hydro-Distillation.

Scientific (common) name	Dry plant mass	Essential oil mass (color)	Percent yield
<i>S. baicalensis</i> (Baikal skullcap)	4.74 g	42.9 mg (pale yellow)	0.9
<i>S. barbata</i> (barbed skullcap) #1 ^a	4.12 g	142.6 mg (colorless)	3.5
<i>S. barbata</i> (barbed skullcap) #2 ^b	2.74 g	145.3 mg (colorless)	5.3
<i>S. incana</i> (hoary skullcap)	8.14 g	155.6 mg (colorless)	1.9
<i>S. lateriflora</i> (blue skullcap)	16.35 g	104.7 mg (pale yellow)	0.6

^a*Scutellaria barbata* #1 sample was harvested on June 7, 2020

^b*Scutellaria barbata* #2 sample was harvested on July 7, 2020.

Flavonoids, diterpenes, alkaloids, glycosides, and volatile essential oil components have all been studied from *Scutellaria* species, with over 295 compounds having been characterized by 2010.³⁷ The presence of neo-clerodane diterpenoid alkaloids with cytotoxic activity in *Scutellaria barbata* has been confirmed experimentally,^{38,39} numerous other neo-clerodane diterpenoids as well as flavonoids,²³ and aromatic terpenoids and hydrocarbons have been analytically revealed. This study examines the volatile composition and enantiomeric distribution of components of the essential oils of these 4 *scutellaria* species and compares these findings with previous literature reports of their essential oil compositions from other geographic locations as well as discusses the relevance of the terpenoids that were found in relation to their ethnobotanical uses.

Results and Discussion

In this study, the essential oils of 4 species of *Scutellaria* most commonly used in traditional medicine were obtained through hydrodistillation and analyzed by gas chromatographic techniques. The following table (Table 1) reports the yields of essential oils obtained from the dried plant matter of the 4 *scutellaria* species.

Table 2, below, compares the essential oil components from the *Scutellaria* spp. as identified by gas chromatographic analysis.

Table 3 lists the enantiomeric distribution percentages for eight of the most prevalent terpenoids found in these *Scutellaria* species. For all the species (−)-linalool was the predominant enantiomer, (+)-(E)-β-caryophyllene was the only enantiomer found in all samples, α-terpineol was found in about a 50/50% split with (−)-α-terpineol being favored by a less than one percent margin. Germacrene D was found only in its (+)-enantiomeric form and it was absent from *S. incana* altogether, δ-cadinene was only found as its (−)-enantiomer and it was not found in *S. incana* or *S. lateriflora*.

Table 2. Essential Oil Chemical Composition of *Scutellaria* Species Cultivated in South Alabama. Retention Indices, RI (CaI), Were Calculated Using a Homologous Series of N-Alkanes on a ZB-5ms Column. RI (Db) Refers to Reference Database Indices.

RI (caI)	RI (db)	Compound	<i>S. baicalensis</i>	<i>S. barbata</i> #1 ^a	<i>S. barbata</i> #2 ^b	<i>S. incana</i>	<i>S. lateriflora</i>
852	853	(3Z)-Hexen-1-ol	-	3.4	2.2	3.7	0.6
867	867	1-Hexanol	-	0.4	0.4	0.5	-
961	959	Benzaldehyde	-	-	-	0.5	2.0
974	973	1-Octen-3-one	-	-	0.5	-	-
977	978	1-Octen-3-ol	6.1	25.6	20.1	31.2	27.5
995	996	3-Octanol	0.6	0.9	0.5	0.5	0.2
1034	1033	Benzyl alcohol	-	-	-	1.7	-
1044	1044	Salicyaldehyde	-	-	-	-	0.6
1064	1064	Acetophenone	-	-	-	1.6	3.6
1098	1099	Linalool	1.2	3.3	2.5	6.8	1.1
1104	1104	Nonanal	-	-	-	1.7	0.3
1112	1111	Phenyl ethyl alcohol	-	-	-	1.0	-
1190	1190	Methyl salicylate	-	-	-	0.2	-
1195	1195	α -Terpineol	0.2	0.3	0.5	0.7	0.1
1197	1197	Methyl chavicol (=Estragole)	-	2.1	-	0.5	-
1215	1217	Coumaran	0.3	1.7	2.3	2.2	0.2
1236	1238	Carvacrol methyl ether	0.8	0.5	1.1	-	-
1238	1238	Neral	-	-	-	0.3	-
1238	1237	Pulegone	-	-	-	-	1.1
1266	1266	Geranial	-	-	-	0.6	-
1285	1290	<i>o</i> -Acetylanisole	-	-	-	-	1.8
1288	1289	Thymol	7.5	2.2	7.7	0.3	2.6
1293	1294	Dihydrodulcan II A	-	-	0.5	1.4	-
1295	1296	Carvacrol	9.3	2.3	8.9	-	0.7
1308	1309	4-Vinylguaiacol	-	-	-	2.1	0.8
1328	1336	δ -Elemene	0.4	-	-	-	-
1350	1356	Eugenol	-	-	-	0.8	0.1
1352	1352	α -Longipinene	-	-	-	-	1.4
1356	1356	(E)-Benzalacetone	-	-	-	-	4.7
1361	1365	Carvacryl acetate	0.2	-	0.4	-	-
1372	1375	α -Copaene	0.2	-	0.2	-	-
1376	1367	Cyclosativene	-	-	-	-	0.2
1377	1379	(E)- β -Damascenone	-	-	-	0.4	-
1378	1380	2- ϕ - β -Funebrene	0.1	-	-	-	-
1381	1382	β -Bourbonene	3.1	1.6	2.0	-	1.0
1386	1387	7- ϕ -Sesquithujene	-	-	-	-	0.4
1389	1390	<i>trans</i> - β -Elemene	0.2	-	-	0.2	0.6

(Continued)

Table 2. Continued

RI (cal)	RI (db)	Compound	S. baicalensis	S. barbata #1 ^a	S. barbata #2 ^b	S. incana	S. lateriflora
1399	1415	Sesquithujene	0.3	-	-	-	-
1402	1405	(Z)- β -Caryophyllene	-	-	-	-	0.5
1412	1412	Longifolene	-	-	-	-	1.0
1418	1417	(D)-β-Caryophyllene	11.6	3.6	2.7	0.6	8.8
1418	1415	α -Barbatene	-	-	-	-	0.2
1426	1430	β -Copaene	0.7	0.4	0.5	-	-
1432	1432	<i>trans</i> - α -Bergamotene	-	-	0.4	-	4.1
1439	1439	(Z)- β -Farnesene	-	-	-	-	0.5
1440	1447	<i>iso</i> -Germacrene D	0.3	-	-	-	-
1450	1452	(E)- β -Farnesene	0.5	-	-	-	0.3
1453	1453	α -Humulene	1.3	0.3	0.6	-	3.2
1452	1447	β -Barbatene	-	-	-	-	0.8
1455	1458	<i>allo</i> -Aromadendrene	0.4	-	-	-	-
1464	1464	<i>allo</i> -Aromadendrene	0.2	-	-	-	-
1471	1475	γ -Murolene	0.2	-	-	-	-
1478	1480	Germacrene D	39.3	11.0	5.4	1.5	
1477	1481	(E)- β -Ionone	-	-	-	0.3	0.1
1480	1482	γ -Himachalene	-	1.1	1.4	-	0.8
1483	1483	<i>trans</i> - β -Bergamotene	0.3	-	-	-	-
1491	1497	Bicyclogermacrene	4.8	1.4	0.6	-	-
1494	1497	α -Murolene	0.3	-	-	-	-
1500	1500	Pentadecane	-	-	-	0.02	-
1501	1503	β -Himachalene	-	-	-	-	0.2
1502	1503	(E,E)- α -Farnesene	0.3	-	-	0.5	-
1506	1509	β -Curcumene	0.7	-	-	-	-
1509	1512	γ -Cadinene	0.3	-	-	-	-
1509	1513	Cuparene	-	-	-	-	0.9
1514	1518	δ -Cadinene	0.8	0.6	1.2	-	-
1522	1523	β -Sesquiphellandrene	-	-	-	-	0.2
1523	1524	Dihydroactinidiolide	-	0.4	0.4	0.6	0.1
1525	1528	(E)- γ -Bisabolene	-	-	-	-	0.1
1535	1535	γ -Cuprenene	-	-	-	-	0.1
1542	1544	<i>di</i> -Sesquibabinene hydrate	-	-	-	-	0.2
1559	1560	(E)-Nerolidol	-	0.4	-	10.5	-
1572	1574	Germacrene D-4-ol	4.3	1.3	1.0	-	-
1577	1577	Caryophyllene oxide	0.5	-	0.4	-	2.4
1589	1594	Viridiflorol	-	0.7	0.6	-	-
1600	1600	Hexadecane	-	0.3	0.5	0.2	0.1

(Continued)

Table 2. Continued

RI (cal)	RI (dlb)	Compound	S. baicalensis	S. barbata #1 ^a	S. barbata #2 ^b	S. incana	S. lateriflora
1612	1613	Humulene epoxide II <i>α</i> -Guai-3,9-dien-11-ol	-	-	-	-	0.4
1634	1639	<i>β</i> -Cadinol	0.2	-	-	-	-
1638	1643	τ-Murrool	0.2	-	-	-	-
1639	1644	<i>α</i> -Cadinol	0.3	-	-	-	-
1651	1655	β-Himachalol	0.8	0.6	0.5	-	-
1657	1652	Patchouli alcohol	-	-	-	-	0.4
1668	1668	(Z)- <i>α</i> -Santalol	-	0.8	-	1.4	-
1670	1674	Germacr-4(15),5,10(14)-trien-1 <i>α</i> -ol	0.2	0.5	-	-	-
1679	1683	Heptadecane	-	-	-	-	-
1700	1700	Octadecane	-	-	-	0.3	0.1
1800	1800	(2Z,6E)-Farnesyl acetate	-	-	-	0.2	-
1827	1832	Neophytadiene	-	-	-	-	0.3
1832	1836	Phytone	-	-	-	-	0.2
1840	1841	Benzyl salicylate	-	-	-	0.4	-
1866	1869	Nonadecane	-	-	0.4	0.2	0.1
1900	1900	Palmitic acid	-	-	13.0	0.5	-
1959	1958	Phytol	1.0	1.8	19.7	14.8	
2107	2102	Linoleic acid	-	2.3	2.3	-	-
2123	2128	Linolenic acid	-	0.7	1.5	-	-
2129	2134	Palmitaldehyde diallyl acetal	-	-	-	0.2	-
2134	2138	Tricosane	-	0.3	0.9	1.2	0.2
2300	2300	Tetracosane	-	-	0.4	-	-
2400	2400	Pentacosane	-	1.0	3.5	1.8	1.0
1500	2500	Hexacosane	-	0.7	0.8	-	-
2600	2600	Heptacosane	-	1.6	5.4	0.8	1.2
2700	2700	Octacosane	-	-	0.9	-	-
2793	2800	Nonacosane	-	-	-	0.7	-
2900	2900	Untriaccontane	-	-	-	0.2	0.2
3100	3100	Total	100.0	99.1	96.9	98.8	98.1

^a*Scutellaria barbata* #1 sample was harvested on June 7, 2020.

^b*Scutellaria barbata* #2 sample was harvested on July 7, 2020.

Table 3. Chirality Data for the Enantiomeric Distribution of Eight of the Most Prevalent Terpenoids in These 4 *Scutellaria* Species Here Examined.

<i>Scutellaria</i> species	Linalool		α -Terpineol		<i>trans</i> - β -Elemene		(E) - β -Caryophyllene	
	% (+)	% (-)	% (+)	% (-)	% (+)	% (-)	% (+)	% (-)
<i>S. baicalensis</i>	29.5	70.5	49.7	50.3	-	-	100	0
<i>S. barbata</i> #1 ^a	16.7	83.3	49.3	50.7	-	-	100	0
<i>S. barbata</i> #2	16.4	83.6	49.9	50.1	-	-	100	0
<i>S. incana</i>	37.3	62.7	49.1	50.9	-	-	100	0
<i>S. lateriflora</i>	18.9	81.1	50.0	50.0	60.3	39.7	100	0
(E) - β -Ionone		(E) -Nerolidol		Germacrene D		δ -Cadinene		
	% (+)	% (-)	% (+)	% (-)	% (+)	% (-)	% (+)	% (-)
<i>S. baicalensis</i>	-	-	-	-	100	0	0	100
<i>S. barbata</i> #1	-	-	-	-	100	0	0	100
<i>S. barbata</i> #2	-	-	-	-	100	0	0	100
<i>S. incana</i>	100	0	4.3	95.7	-	-	-	-
<i>S. lateriflora</i>	-	-	-	-	100	0	-	-

^a*Scutellaria barbata* #1 sample was harvested on June 7, 2020.

^b*Scutellaria barbata* #2 sample was harvested on July 7, 2020.

The volatile phytochemical makeup of the essential oils of these 4 *scutellaria* species were collectively dominated by 1-octen-3-ol, thymol, carvacrol, (E) - β -caryophyllene, germacrene D, (E) -nerolidol, palmitic acid, phytol, linalool, β -bourbonene, and linolenic acid. Studies of the essential oil from *S. barbata* from the Hunan region of China showed a predominance of 1-octen-3-ol, hexadecanoic acid (palmitic acid), and (E) - β -caryophyllene,⁴⁰ however most of the other key aromatic molecules such germacrene D, carvacrol, thymol, phytol, and linalool found in our research were not reported in their analysis. A previous study from the University of Wuhan, China showed the essential oil of *S. barbata* to exhibit more similar molecular makeup to our samples, with hexahydrofarnesylacetone, 3,7,11,15-tetramethyl-2-hexadecen-1-ol, menthol, and 1-octen-3-ol being the dominant by percentage, but also in significant quantities (>1%) were linalool, β -bourbonene, β -himachalene, and thymol.⁴¹

In their 2010 review of several *Scutellaria* essential oil studies, Shang et al.,³⁷ noted high concentrations of (E) - β -caryophyllene and linalool. Samples of essential oils from wild-harvested *Scutellaria* species from Uzbekistan also contained linalool, (E) - β -caryophyllene, thymol, and germacrene D,⁴² similar to our findings of these volatile components in the essential oils of Alabama-grown *Scutellaria*.

The aromatic terpenoids of herbs and flowers have a long history of aromatherapeutic and internal uses as teas and decoctions to treat an extremely wide range of disease states. Terpenes have been shown to be effective antibacterial, antifungal, anti-viral, anti-inflammatory, neuro-protectives, anti-tumor and anti-proliferative as well as uses for dental and dermatological treatments and other uses⁴³ including bacteriostatic surface cleaners⁴⁴ and pesticides,⁴⁵ in the case of thymol. Interestingly, 1-octen-3-ol, the most

prevalent aromatic compound in all except one out of these four essential oils (*S. baicalensis*) is known for being a chemical feeding attractant for certain varieties of mosquito while acting as a repellent to other strains of mosquito.⁴⁶ It is also known as the “mushroom acid,” and is the most prevalent aromatic compound in most mushrooms (including truffles), molds, and fungi and is the primary olfactory irritant in rotting wood. Over-exposure to this aromatic compound has been shown to cause conjunctivitis, rhinitis, and respiratory inflammation.⁴⁷ This compound is also found throughout nature, in raspberries, elder flowers, in the molded rind of Camembert cheese, in certain unwanted wine-molds, and is expressed as a pheromone by certain millipedes during distress.⁴⁸ The two enantiomers of this compound are known to have different aromas and may exhibit differing effects when it comes to inflammatory response. Furthermore, 1-octen-3-ol has shown antibacterial and anti-fungal activities.⁴⁹

In this case it is significant to note the finding of a significant percentage of phytol in both *S. lateriflora* and *S. incana*, the American herbs, but not the Asian species. Phytol has shown promise for controlling the neglected tropical disease schistosomiasis^{50,51} as well as showing antibacterial properties against *Staphylococcus aureus*⁵² and *Mycobacterium tuberculosis*.⁵³ Phytol has also been shown to be an anti-allergen and when applied topically in murine studies.⁵⁴ Antioxidant and antinociceptive properties of phytol have also been observed.⁵⁵

A high percentage (>5%) of (E) -nerolidol and linalool was unique to *S. incana*. Studies of Greek varieties of *Scutellaria* also showed linalool and nerolidol to be the main constituents of the essential oil and testing showed moderate antimicrobial activity.⁵⁶ (E) -Nerolidol has shown low toxicity with unspecified increased respiratory function



Figure 1. *Scutellaria baicalensis* photographed at Kirkland Gardens just before harvest. Photograph by S. K. Lawson, June 2020.



Figure 2. *Scutellaria barbata*, photographed in May, 2020, by Kirk Lawson.

as well as cytotoxic effects again hepatic carcinomas.⁵⁷ Plant essential oils with high percentages of nerolidol have been found to kill *Schistosoma*⁵⁸ and have antileishmanial activity.⁵⁹ (E)-Nerolidol has shown promise as an antifungal in guinea pig testing by healing fungal skin lesions,⁶⁰ and as an antiulcer agent as well.⁶¹ Linalool has been experimentally shown to be antimicrobial, cytotoxic, antioxidant, anti-inflammatory, and have insect repellent properties while showing almost no toxicity or allergic response and near zero skin irritant traits.⁶²

Linalool also has been shown in-vivo to have significant sedative⁶³ and anti-convulsant effects⁶⁴ and is one of the main aromatic compounds in lavender essential oils.⁶⁵ Linalool has been found to be a potent acetylcholinesterase inhibitor and has potential as an Alzheimer's therapeutic essential oil.⁶⁶

The native Asian species showed relatively high percentages of carvacrol and thymol where the native American species did not. The largest percentage of any single compound in the essential oils examined herein was the presence of germacrene D at 39% in *S. baicalensis*. *Scutellaria baicalensis* essential oil from the aerial parts in this study was rich in (+)-germacrene D (39.3%), (+)-(E)- β -caryophyllene (11.6%), carvacrol (9.3%), thymol (7.5%), and 1-octen-3-ol (6.1%). A headspace volatile analysis of *S. baicalensis* flowers showed the floral volatiles to be dominated by sesquiterpene hydrocarbons, particularly (E)- β -caryophyllene (22.3 to 41.5%) and germacrene D (12.4 to 27.5%), but carvacrol, thymol, or 1-octen-3-ol were not reported.⁶⁷ The root

essential oil of *S. baicalensis* was reported, but the constituents were not quantified.⁶⁸ Based on the gas chromatogram, the root oil had acetophenone, (E)-4-phenyl-2-butanone, palmitic acid, and oleic acid as major components; (E)- β -caryophyllene was detected, but in low concentration; and neither carvacrol nor thymol were detected. The analgesic and anti-inflammatory properties of (E)- β -Caryophyllene have been shown experimentally.⁶⁹⁻⁷⁴ Both thymol and carvacrol has shown notable antimicrobial activities against pathogenic microorganisms⁷⁵⁻⁷⁸ and thymol has shown a wide array of pharmacological properties.⁷⁹ The high concentrations of these compounds may contribute to the biological activities and traditional uses of *S. baicalensis*.

As far as we are aware, this study is the first to report the essential oil composition of *S. incana*. The aerial parts essential oil was rich in 1-octen-3-ol (31.2%), linalool (6.8%), (E)-nerolidol (10.5%), and phytol (19.7%). The (−)-enantiomers predominated for both linalool (63%) and (E)-nerolidol (96%). The essential oil of *S. lateriflora* cultivated in Alabama was also rich in 1-octen-3-ol (27.5%) and phytol (14.8%) in addition to (E)- β -caryophyllene (8.8%). The chemical composition of *S. lateriflora* cultivated in Alabama is in marked contrast to the composition reported for *S. lateriflora* from Iran, which was dominated by sesquiterpenes (78.3%) including τ -cadinene (27.0%), calamenene (15.2%), and β -elemene (9.2%).⁸⁰ *Scutellaria lateriflora* has been shown in rat experiments to have anticonvulsant capabilities in its flavone



Figure 3. *Scutellaria incana*. Photograph taken by S. K. Lawson immediately prior to harvest in June 2020.

component extract,⁸¹ and linalool, found in all four of these *Scutellaria* species, has been shown to be a sedative and anti-convulsant in murine experiments,^{63,64} thus supporting and explaining the pharmacology of the traditional use of this plant for such purposes.

Conclusions

This horticulture, distillation, GC-MS, GC-FID, and chirality analysis of the popular Asian medicinals, *S. barbata* and *S. baicalensis*, as well as the native *Scutellaria* species to Alabama and the southeastern United States, *S. incana* and *S. lateriflora*, aims to better characterize the expected volatile composition of these *Scutellaria* species. With the ability to compare the terpenoid distributions one can make a more well-informed decision as to the true identity of commercially available essential oils. This will also help users of phytotherapies to discern the relevance of the traditional Asian medicine and Native American medicinal claims for the uses of these herbs for inhalation therapies, baths, and teas which can, in certain instances, be supported simply by the appearance of certain aromatic terpenes like thymol, linalool, nerolidol, and β -caryophyllene, with their experimentally supported wide-ranging medicinal uses.



Figure 4. *Scutellaria lateriflora*. Photographed by S. K. Lawson immediately prior to harvest in June 2020.

Experimental

Plant Material

Scutellaria baicalensis seeds (www.Amazon.com—Outsidepride seed store), *Scutellaria barbata* seeds (Plants with a Purpose — www.plantswithapurpose.net), *Scutellaria incana* seedlings (Prairie Moon Nursery—www.prairiemoon.com), and *Scutellaria lateriflora* seeds (Palm Beach Medicinals—www.palmbeachherbs.com) were grown by Kirkland Gardens LLC., in Newville, Alabama. The seeds were germinated in trays of organic peat moss outdoors in March 2020, then when approximately six inches tall they, along with the *S. incana* seedlings, were transplanted to a shady location in the garden under a tree with partial sunlight. The plants were cultivated either in one- or two-gallon black nursery pots with a mix of peat moss, perlite, and cow manure growing medium or in the ground with the same amendments/growing media. The plants were fertilized with chicken manure, bone meal, and kelp meal during transplanting and then watered regularly with well water by drip emitter system. Plants were harvested mid-flowering period, which was the week of June 7, 2020. A second harvesting of *S. barbata* occurred July 7, 2020. The aerial parts of the plants were photographed then cut at ground level and identifications verified with comparison to online plant databases^{24,26-32} and *S. incana* and *S. lateriflora* were positively identified by Kirk Lawson using the Manual of the Vascular

Flora of the Carolinas.⁸² The plant material was allowed to dry indoors in an air-conditioned area at approximately 23 °C for 10 days. Then, the dried aerial parts were vacuum-sealed and shipped to the University of Alabama in Huntsville. Over the weeks of July 24–August 26, 2020, they were and hydro-distilled with a Likens-Nickerson apparatus with continuous extraction with dichloromethane (DCM) for approximately 4 hours each. The DCM extracts were collected, and the small amount of solvent evaporated under a flow hood. The essential oils (EO's) were then stored at –20 °C until further analysis.

Photographs of the plants during flowering and before collection were taken by the author, Kirk Lawson (Figures 1–4). These photographs are stored for reference on multiple hard drives including all of the authors of this manuscript and the digital herbarium at the University of Alabama in Huntsville.

Gas Chromatography

The essential oils were analyzed by gas chromatography–mass spectrometry (GC-MS), gas chromatography with flame ionization detection (GC-FID), and chiral GC-MS as previously reported.⁸³ For GC-MS the Shimadzu GCMS-QP2010 Ultra was utilized, with a ZB-5ms GC column, GC oven temperature of 50 °C – 260 °C (2 °C/min), 1 µL injection of 5% solution of EO in dichloromethane (split mode, 30:1). The retention indices (RIs) were determined by using a series of reference *n*-alkanes. The compounds listed in Table 2 were identified by comparing the mass spectrometry data and retention indices with those in the databases.^{84–87} Gas Chromatography–Flame Ionization Detection was conducted using a Shimadzu GC 2010 with FID detector, ZB-5 GC column, GC oven temperature 50 °C– 260 °C (2 °C/min). The percent compositions were determined from raw peak areas without standardization. Chiral Gas Chromatography–Mass Spectrometry was carried out with a Shimadzu GCMS-QP2010S. A Restek B-Dex 325 column was utilized with an initial GC oven temperature of 50 °C, which was increased gradually to 120 °C at a rate of 1.5 °C/minute. Then the temperature was increased to 200 °C at the rate of 2.0 °C/minute. Exactly 0.1 µL injection of 5% solution of essential oil samples in dichloromethane (split mode, 45:1) was injected. The enantiomeric distributions were determined by comparison of retention times with authentic samples obtained from Sigma-Aldrich (Milwaukee, WI, USA). The relative enantiomer percentages were calculated from peak areas.

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